Product Data Sheet

APC anti-BrdU

Catalog # / 2420565 / 25 tests

Size: 2420570 / 100 tests

Clone: 3D4

Isotype: Mouse IgG1, ĸ

Immunogen: Iodouridine-conjugated ovalbumin

Preparation: The antibody was purified by affinity

> chromatography and conjugated with APC under optimal conditions. The solution is free of unconjugated APC

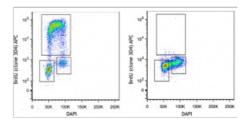
and unconjugated antibody.

Formulation: Phosphate-buffered solution, pH 7.2,

containing 0.09% sodium azide and

0.2% (w/v) BSA (origin USA).

Concentration: Lot-specific



Ramos cell line was pulsed with BrdU for 1 hour (left panel) or without (right panel) and then stained with anti-BrdU (3D4) APC. Cells were subsequently stained with 1 µg of DAPI for DNA

analysis.

Applications:

Applications: Intracellular Flow Cytometry

Recommended

Usage:

Notes:

Each lot of this antibody is quality control tested by intracellular immunofluorescent staining with flow cytometric analysis. For flow cytometric staining, the suggested use of this reagent is 5 µl per million

cells in 100 µl staining volume or 5 µl per 100 µl of whole blood.

Application

Additional reported applications (for the relevant formats) include:

immunohistochemistry and fluorescence microscopy.

Application References:

1. Dolbeare F, et al. 1983. Proc. Natl. Acad. Sci. USA 80:5573.

2. Hirota K, et al. 2007. J. Exp. Med. 204:41.

3. Godebu E, et al. 2008. J. Immunol. 181:1798.

4. Waskow C, et al. 2008. Nat. Immunol. 9:676.

BrdU is a uridine derivative and a structural analog of thymidine, which can **Description:**

be incorporated into DNA during the S-phase of a cell cycle as a substitute for thymidine. Cells can be pulse-labeled with BrdU and analyzed with antibodies against BrdU to determine the proportion of cells in the S-phase

of the cell cycle during a given interval.

Antigen References: 1. Barker JM, et al. 2013. PLoS One 8:e63692.

2. Duque A and Rakic P. 2011. J. Neurosci. 31:15205.

3. Robbins S, et al. 2011. J. Vis. Exp. 55:2855.

4. Broekhuizen CA, et al. 2010. Infect Immun. 78:954.

5. van der Wath RC, et al. 2009. PLoS One 4:e6972.

6. Dolbeare F, et al. 1985. Cytometry 6:521.

7. Gratzner HG. 1982. Science 218:474.