### **Brilliant Violet 605™ anti-human CD11c**

**Catalog # / Size:** 2108180 / 100 tests

2108175 / 25 tests

**Clone:** 3.9

**Isotype:** Mouse IgG1, κ

Reactivity: Human

**Preparation:** The antibody was purified by affinity

chromatography and conjugated with Brilliant Violet 605<sup>™</sup> under optimal conditions. The solution is free of unconjugated Brilliant Violet 605<sup>™</sup> and

unconjugated antibody.

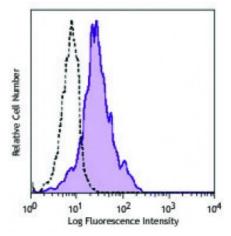
**Formulation:** Phosphate-buffered solution, pH 7.2,

containing 0.09% sodium azide and BSA

(origin USA).

Workshop Number: III NL707

Concentration: Lot-specific



Human peripheral blood granulocytes were stained with CD11c (clone 3.9) Brilliant Violet 605™ (filled histogram) or mouse IgG1, κ Brilliant Violet 605™ isotype control (open histogram).

#### **Applications:**

**Applications:** Flow Cytometry

Recommended

Usage:

Each lot of this antibody is quality control tested by immunofluorescent staining with flow cytometric analysis. For flow cytometric staining, the suggested use of this reagent is ≤5 microL per million cells or 5 microL per 100 microL of whole blood. It is recommended that the reagent be titrated for optimal performance for each application.

Brilliant Violet  $605^{\text{TM}}$  excites at 405 nm and emits at 603 nm. The bandpass filter 610/20 nm is recommended for detection, although filter optimization may be required depending on other fluorophores used. **Be sure to verify that your cytometer configuration and software setup are appropriate for detecting this channel.** Refer to your instrument manual or manufacturer for support. Brilliant Violet  $605^{\text{TM}}$  is a trademark of Sirigen Group Ltd.

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Application Notes:

Clone 3.9 preferentially binds the activated form of CD11c, is specific for the I domain of CD11c, and is able to partially block the binding of CD11c and ICAM-4. 3.9 binding is divalent cation dependent<sup>12</sup>. While analyzing blood, it is best to use heparin as the anti-coagulant and not EDTA. Since the ability of clone 3.9 to bind to its target is divalent cation dependent, the usage of EDTA as an anti-coagulant may be detrimental to staining due to its chelating properties.

Additional reported applications (for the relevant formats) include: immunohistochemical staining of acetone-fixed frozen tissue sections4, and functional assays<sup>5,6</sup>. The LEAF<sup>m</sup> purified antibody (Endotoxin <0.1 EU/ $\mu$ g, Azide-

Free, 0.2  $\mu$ m filtered) is recommended for functional assays (Cat. No. 301616). For highly sensitive assays, we recommend Ultra-LEAF<sup>™</sup> purified antibody (Cat. No. 301632) with a lower endotoxin limit than standard LEAF<sup>™</sup> purified antibodies (Endotoxin <0.01 EU/microg).

# Application References:

- 1. Schlossman S, *et al.* Eds. 1995. Leucocyte Typing V. Oxford University Press. New York.
- 2. Knapp W, et al. 1989. Leucocyte Typing IV Oxford University Press. New York.
- 3. McMichael A, et al. Eds. 1987. Leucocyte Typing III Oxford University Press. New York
- 4. Vainer B, et al. 2000. Am. J. Surg. Pathol. 24:1115. (IHC)
- 5. Ottonello L, et al. 1999. Blood 93:3505.
- 6. Metelitsa LS, et al. 2002. Blood 99:4166.
- 7. Sadhu C, et al. 2007. J. Leukoc. Biol. doi:10.1189/jlb.1106680. PubMed
- 8. Ihanus E, et al. 2007. Blood 109:802-810.
- 9. Gurer C, et al. 2008. Blood 112:1231. PubMed
- 10. Asai A, et al. 2009. J. Lipid Res. 50:95. PubMed
- 11. Yoshino N, et al. 2000. Exp. Anim. (Tokyo) 49:97. (FC)
- 12. Sadhu C, et al. 2008. J. Immunoass. Immunoch. 29:42. (FC)

#### **Description:**

CD11c is a 145-150 kD type I transmembrane glycoprotein also known as integrin  $\alpha_X$  and CR4. CD11c non-covalently associates with integrin  $\beta$ 2 (CD18) and is expressed on monocytes/macrophages, dendritic cells, granulocytes, NK cells, and subsets of T and B cells. CD11c has been reported to play a role in adhesion and CTL killing through its interactions with fibrinogen, CD54, and iC3b.

## Antigen References:

- 1. Petty H. 1996. Immunol. Today 17:209.
- 2. Springer T. 1994. Cell 76:301.
- 3. Ihanus E, et al. 2007. Blood 109:802-810.