

APC anti-mouse CD135

Catalog # / Size: 1276545 / 25 µg
1276550 / 100 µg

Clone: A2F10

Isotype: Rat IgG2a, κ

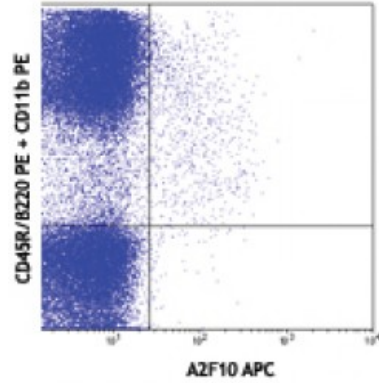
Immunogen: Mouse Flt3 transfected cell line

Reactivity: Mouse

Preparation: The antibody was purified by affinity chromatography, and conjugated with APC under optimal conditions. The solution is free of unconjugated APC and unconjugated antibody.

Formulation: Phosphate-buffered solution, pH 7.2, containing 0.09% sodium azide.

Concentration: 0.2

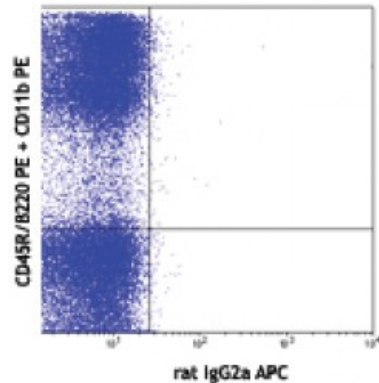


C57BL/6 bone marrow cells stained with A2F10 APC and CD45R/B220 (RA3-6B2) plus CD11b (M1/70) PE

Applications:

Applications: Flow Cytometry

Recommended Usage: Each lot of this antibody is quality control tested by immunofluorescent staining with flow cytometric analysis. For flow cytometric staining, the suggested use of this reagent is ≤ 1.0 microg per 10⁶ cells in 100 microL volume. It is recommended that the reagent be titrated for optimal performance for each application.



C57BL/6 bone marrow cells stained with rat IgG2a APC isotype control and CD45R/B220 (RA3-6B2) plus CD11b (M1/70) PE

- Application References:**
1. Sergejeva S, *et al.* 2004. *Blood* 103:1270.
 2. Auffray C, *et al.* 2009. *J. Exp. Med.* 206:595.

Description: CD135, also known as Flk-2, Flt3, and Ly-72, is a type III tyrosine kinase receptor. It is expressed on early B lymphoid lineage cells in bone marrow, on primitive myeloid progenitors within the BM CD34+ cell population. Ligation of Flk-2 with Flt3 ligand regulates the growth of hematopoietic stem cells and promotes the survival of primitive hematopoietic progenitor cells with myeloid as well as B lymphoid potential. It was reported that the receptor tyrosine kinase Flt3 is required for dendritic cell development. Combined signaling through interleukin-7 receptors and Flt3 selectively promotes B-cell commitment and differentiation from uncommitted murine bone marrow progenitor cells.

- Antigen References:**
1. Waskow C, *et al.* *Nat. Immunol.* 9:676
 2. Veiby OP, *et al.* 1996. *Blood* 88(4):1256
 3. Veiby OP, *et al.* 1996. *J. Immunol.* 157(7):2953
 4. Matthews W, *et al.* 1991. *Cell.* 65(7):114

